



## Product Specifications

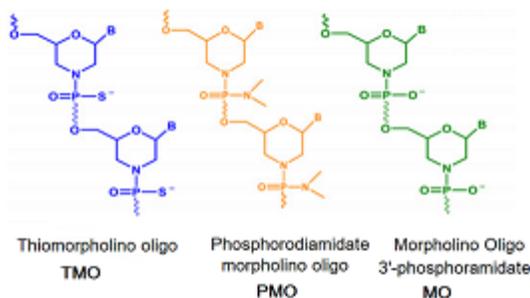
Custom Oligo Synthesis, antisense oligos, RNA oligos, chimeric oligos, Fluorescent dyes, Affinity Ligands, Spacers & Linkers, Duplex Stabilizers, Minor bases, labeled oligos, Molecular Beacons, siRNA, phosphonates Locked Nucleic Acids (LNA); 2'-5' linked Oligos

## Oligo Modifications

For research use only. Not for use in diagnostic procedures for clinical purposes.

### Morpholino Oligo G [MO-G]

Category	Antisense
Modification Code	MO-G
Reference Catalog Number	26-6644G
5 Prime	Y
3 Prime	Y
Internal	Y
Molecular Weight(mw)	328.29



Thio morpholino oligos modification has a setup charge of \$250.00 per order for special synthesis reagents.

**SPECIAL NOTE:** Please note below the maximum number of sites that can be incorporated to achieve a reliable yield of the modified oligo.

**TMO Sites.** The maximum number of thiomorpholino oligo(TMO) sites is 8 per oligo. These sites can be spread out in the sequence and the preferred construct is (4+4); 4 sites at the 5' end and 4 sites at the 3' end.

**MO Sites.** The maximum number of MO sites is 8 per oligo. These sites can be spread out in the sequence and the preferred construct is (4+4); 4 sites at the 5' end and 4 sites at the 3' end.

#### Morpholino Oligos (PMO)

Phosphorodiamidate morpholino oligos (PMOs) are chemically modified oligonucleotides antisense oligonucleotides (ODN, ASO) wherein the 2'-deoxyribonucleosides and phosphate linkages of canonical DNA are substituted with morpholino rings and phosphorodiamidate linkages respectively. By replacing the standard ribose or deoxyribose sugar and the phosphodiester linkages with six-membered morpholino phosphorodiamidate linkages the anionic linkages become non-ionic. These morpholinos rings may provide higher conformational rigidity when incorporated into an oligonucleotide backbone

They have been researched extensively in oligonucleotide therapeutics as potential steric blocking agents for the treatment of various genetic disorders. PMOs exhibit high hybridization affinity to complementary RNA, possess excellent enzymatic stability both in vitro and in vivo and elicit low immunogenicity leading to acceptable toxicokinetic profiles in mammalian models. The ability of PMOs to effectively function as steric blockers arises from their ability to specifically bind complementary RNA both in vitro and in vivo. Morpholinos block small (~25 base) regions of the base-pairing surfaces of ribonucleic acid (RNA).

Morpholino oligos are specific, soluble, non-toxic, stable, and effective antisense oligonucleotide suitable for development as therapeutics and currently in clinical trials. Efficacy of Morpholino oligos in humans has been shown in clinical trials for Duchenne muscular dystrophy. The splice modifying Morpholino eteplisen has partially restored function to the dystrophin protein, enough to show significant clinical benefit on a six-minute walk test versus the untreated control group.

Blocking translation- Morpholinos can bind to the 5'-untranslated region of messenger RNA (mRNA) and can interfere with progression of the ribosomal initiation complex from the 5' cap to the start codon.

Modifying pre-mRNA splicing- Morpholinos can prevent splice-directing small nuclear ribonucleoproteins (snRNP) complexes from binding introns of pre-mRNA, blocking the splice lariat structure, interfering with the binding of splice regulatory proteins such as splice silencers. [1] and splice enhancers [2] there by interfering with pre-mRNA processing steps.

Morpholinos can block miRNA activity [3][4] and maturation.[5] Fluorescein-tagged Morpholinos along with fluorescein-specific anti-bodies that could be used as probes for in-situ hybridization to miRNAs [6] Morpholinos can also block ribozyme activity. [7]

Gene knockdown by morpholino- The morpholino-modified antisense oligonucleotides are primarily used in animal embryonic systems for the 'knock-down' of gene function Intron retention using RNA-targeting thiomorpholino antisense oligonucleotides.[8]

### **Morpholino Oligos References**

1. Stirchak, E. P., Summerton, J. E., & Weller, D. D. (1989). Uncharged stereoregular nucleic acid analogs: 2. Morpholino nucleoside oligomers with carbamate internucleoside linkages. *Nucleic acids research*, 17(15), 6129-6141. <https://doi.org/10.1093/nar/17.15.6129>
2. Krishna, Heera; Jastrzebska, Katarzyna; Caruthers, Marvin, *FEBS Letters*, 17 June 2019, Volume593, Issue13 ,Krakow Special Issue Pages 1459-1467, <https://doi.org/10.1002/1873-3468.13492>
3. Bruno IG, Jin W, Cote GJ (October 2004). Correction of aberrant FGFR1 alternative RNA splicing through targeting of intronic regulatory elements. *Human Molecular Genetics*. 13(20): 2409-20. doi:10.1093/hmg/ddh272. PMID 15333583.
4. Vetrini F, Tammara R, Bondanza S, Surace EM, Auricchio A, De Luca M, Ballabio A, Marigo V (May 2006). Aberrant splicing in the ocular albinism type 1 gene (OA1/ GPR143) is corrected in vitro by morpholino antisense oligonucleotides. *Human Mutation*. 27 (5): 420-6. doi:10.1002/humu.20303. PMID 16550551.
5. Kloosterman WP, Wienholds E, Ketting RF, Plasterk RH (2004). Substrate requirements for let-7 function in the developing zebrafish embryo. *Nucleic Acids Research*. 32 (21): 6284-91. doi:10.1093/nar/gkh968. PMC 535676. PMID 15585662.
6. Flynt AS, Li N, Thatcher EJ, Solnica-Krezel L, Patton JG (February 2007). Zebrafish miR-214 modulates Hedgehog signaling to specify muscle cell fate. *Nature Genetics*. 39 (2): 259-63. doi:10.1038/ng1953. PMC 3982799. PMID 17220889.
7. Kloosterman WP, Lagendijk AK, Ketting RF, Moulton JD, Plasterk RH (August 2007). Targeted inhibition of miRNA maturation with morpholinos reveals a role for miR-375 in pancreatic islet development. *PLOS Biology*. 5 (8): e203. doi:10.1371/journal.pbio.0050203. PMC 1925136. PMID 17676975
8. Lagendijk AK, Moulton JD, Bakkens J (June 2012). Revealing details: whole mount microRNA in situ hybridization protocol for zebrafish embryos and adult tissues. *Biology Open*. 1 (6): 566-9. doi:10.1242/bio.2012810. PMC 3509442. PMID 23213449.
9. Yen L, Svendsen J, Lee JS, Gray JT, Magnier M, Baba T, D'Amato RJ, Mulligan RC (September 2004). Exogenous control of mammalian gene expression through modulation of RNA self-cleavage. *Nature*. 431 (7007): 471-6. Bibcode:2004Natur.431..471Y. doi:10.1038/nature02844. PMID 15386015.
10. D. Gabrijela, B. Ulrich, K.Heera, J. Katarzyna, S. Michael, B. Benjamin, C.Thomas, C.Marvin, R.John Nuclear compartmentalization of TERT mRNA and TUG1 lncRNA transcripts is driven by intron retention: implications for RNA-directed therapies.
11. K.Heera, J. Katarzyna, C. Marvin, P. Sibasish, V.Rakesh WO2019060522 - THIOMORPHOLINO OLIGONUCLEOTIDES FOR THE TREATMENT OF MUSCULAR DYSTROPHY.
12. Guide for Morpholino Users: Toward Therapeutics *J Drug Discov Develop and Deliv*. 2016; 3(2): 1023. Gene Tools, LLC, USA \*Corresponding author: Moulton JD, Gene Tools, LLC, 1001 Summerton Way, Philomath, Oregon 97370, USA.