

# Product Specifications

Custom Oligo Synthesis, antisense oligos, RNA oligos, chimeric oligos, Fluorescent dyes, Affinity Ligands, Spacers & Linkers, Duplex Stabilizers, Minor bases, labeled oligos, Molecular Beacons, siRNA, phosphonates Locked Nucleic Acids (LNA); 2'-5' linked Oligos

## Oligo Modifications

For research use only. Not for use in diagnostic procedures for clinical purposes.

### L-RNA rG

Category	Others	<b>O</b>	
Modification Code	L-rG	HN	
Reference Catalog Number	27-6943	H <sub>2</sub> N N N FO	0-^^ 5' Oligo
5 Prime	Υ	0_0	—P=0
3 Prime	Υ		OH
Internal	Υ	OH O	L-RNA rG [27-6943-XX]
Molecular Weight(mw)	345.21	Oligo 3'	
		ÓН	

L-RNA (beta stereoisomer ribose that is same as in D-RNA) is the left-turning and mirror image version of natural RNA, as opposed to the naturally occurring right-turning version called D-RNA. L-RNA is more stable than D-RNA to enzymatic degradation by certain nucleases (1). Since the two enantiomers are identical in structure other than their chiral differences, their intrinsic physical properties are generally equal to each other. This includes duplex stability, solubility, and selectivity as D-RNA but form a left-helical double-helix. Because of its chiral difference, L-RNA does not bind to its naturally occurring D-RNA counterpart.

One important aspect of L-RNA is that it is poor at hybridizing to D-RNA (2). This confers multiple uses, one being that the incorporation of L-RNA into the stem of a molecular beacon as it allows stem invasion to be avoided (3). Other areas that it can play an important role in would be zip-code microarrays (2) and as molecular tags for PCR (4). When used in nanocarriers, L-DNA has greater cellular uptake as well as greater serum stability. It is good for also reducing interaction between aptamers and nanocarrier skeletons (5).

Gene Link synthesizes L-RNA oligos with any combination of D-RNA bases including fluorescent dyes and all other available modifications and chimeric bases.

#### L-DNA Applications

#### - References

- 1) Damha M.J., Giannaris P.A., Marfey P. Antisense L/D-oligonucleotide chimeras: nuclease stability, base-pairing properties, and activity at directing ribonuclease H. Biochemistry. 1994;33:7877?7885.<
- 2) Utilising the left-helical conformation of L-DNA for analysing different marker types on a single universal microarray platform Nicole C. Hauser, Rafael Martinez, Anette Jacob, Steffen Rupp, J"rg D. Hoheisel, Stefan Matysiak Nucleic Acids Res. 2006 October; 34(18): 5101-5111. Published online 2006 September 20. doi:ÿ10.1093/nar/gkl671
- 3) Superior structure stability and selectivity of hairpin nucleic acid probes with an L-DNA stem. Youngmi Kim, Chaoyong James Yang, Weihong Tan Nucleic Acids Res. 2007 December; 35(21): 7279?7287. Published online 2007 October 24. doi:ÿ10.1093/nar/gkm771
- 4) Hayashi G., Hagihara M., Nakatani K. Application of L-DNA as a molecular tag. Nucleic Acids Symp. Ser. 2005;49:261?262
- 5) Utilizing the bioorthogonal base-pairing system of L-DNA to design ideal DNA nanocarriers for enhanced delivery of nucleic acid cargos Kyoung-Ran Kim, Taemin Lee, Byeong-Su Kim and Dae-Ro Ahn.



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